

**REFERENCES**

1. Urine Testing for Drugs of Abuse, National Institute on Drug Abuse Research Monograph, 73,1986.
2. R.C. Baselt. In : Advances in Analytical Technology, Vol.1. Randall C. Baselt edd. (Biomedical Publications, Foster City, CA. 87-93).
3. Driscoll, R.C., Barr, F.S., Gragg, B.J. and G.W. Moore. Determination of Therapeutic Blood Levels of Methamphetamine by GC. J.Pharm. Sci 60:1492.1971.

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For Research Use Only. Not for use in Diagnostic Procedures.

**Methamphetamine ELISA**

Catalog No. ME090D

**INTENDED USE**

The Calbiotech Methamphetamine Direct ELISA Kit is a specific and sensitive in-vitro test to detect the presence of d-methamphetamine in samples such as whole blood, oral fluids, serum, plasma and urine.

**SUMMARY AND EXPLANATION**

While the assay will detect amphetamine use, interference by l-methamphetamine and pseudoephedrine is virtually nonexistent. Methamphetamine is a potent central nervous system stimulant with less peripheral actions than mphetamine. The (+)-isomer also referred to as d-methamphetamine is ten times more potent than the (-)-isomer, l-methamphetamine. Amphetamines act by inducing euphoria, irritability, anxiety and paranoia Methamphetamine is metabolized to its active metabolite amphetamine (via N-demethylation) and is further metabolized by hydroxylation and deamination of amphetamine. Urinary excretion rates are influenced by the urinary pH with acidic urine favoring the excretion of unchanged drug. Alkaline urine reduces the excretion of unchanged methamphetamine to less than 5% of the dose.

**PRINCIPLES OF THE TEST**

The Methamphetamine Direct ELISA Kit (for d-methamphetamine measurement) is based upon the competitive binding to antibody of enzyme labeled antigen and unlabeled antigen, in proportion to their concentration in the reaction mixture. A 10 µl. aliquot of a diluted unknown specimen is incubated with a 100 µl. dilution of enzyme (Horseradish peroxidase) labeled d-methamphetamine derivative in micro-plate wells, coated with fixed amounts of oriented high affinity purified polyclonal antibody. The wells are washed thoroughly and a chromogenic substrate added. The color produced is stopped using a dilute acid stop solution and the wells read at 450 nm. The intensity of the color developed is inversely proportional to the concentration of drug in the sample. The technique is sensitive to 1 ng/ml. The Methamphetamine Direct ELISA Kit avoids extraction of urine sample for measurement. It employs a d-methamphetamine directed antiserum. Due to the proprietary method of orienting the antibody on the polystyrene micro-plate much higher sensitivity is achieved compared to passive adsorption. This allows an extremely small sample size, reducing matrix effects and interference with binding proteins(s) or other macromolecules.

Cat#: ME090D  
For Order and Inquiries, please contact  
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MATERIAL PROVIDED	96Tests
1. Microwells coated with polyclonal anti-d-methamphetamine	12x8x1
2. d-Methamphetamine Conjugate	12.5 ml
3. Positive Reference Standard	1ml
4. Negative Standard	1ml
5. TMB Substrate	14 ml
6. Stop Solution	12.5 ml

**MATERIALS NOT PROVIDED**

1. Distilled or deionized water.
2. Precision pipettes. Disposable pipette tips.
3. ELISA reader capable of reading absorbance at 450nm.
4. Absorbance paper or paper towel.
5. Graph paper.

**STORAGE AND STABILITY**

1. Store the kit at 2 - 8° C.
2. Keep microwells sealed in a dry bag with desiccants.
3. The reagents are stable until expiration of the kit.
4. Do not expose test reagents to heat, sun, or strong light.

**WARNINGS AND PRECAUTIONS**

1. Potential biohazardous materials:  
The calibrator and controls contain human source components which have been tested and found non-reactive for hepatitis B surface antigen as well as HIV antibody with FDA licensed reagents. However, as there is no test method that can offer complete assurance that HIV, Hepatitis B virus or other infectious agents are absent, these reagents should be handled at the Biosafety Level 2, as recommended in the Centers for Disease Control/National Institutes of Health manual, "Biosafety in Microbiological and Biomedical Laboratories." 1984
2. This test kit is designed for Research Use Only. Not for use in diagnostic procedures.
3. Do not pipette by mouth. Do not smoke, eat, or drink in the areas in which specimens or kit reagents are handled.
4. The components in this kit are intended for use as an integral unit. The components of different lots should not be mixed.
5. It is recommended that serum samples be run in duplicate.
6. Optimal results will be obtained by strict adherence to this protocol. Accurate and precise pipetting, as well as following the exact time and temperature requirements prescribed are essential. Any deviation from this may yield invalid data.

**SPECIMEN COLLECTION HANDLING**

1. Collect blood specimens and separate the serum immediately.
2. Specimens may be stored refrigerated at (2-8°C) for 5 days. If storage time exceeds 5 days, store frozen at (-20° C) for up to one month.
3. Avoid multiple freeze-thaw cycles.
4. Prior to assay, frozen sera should be completely thawed and mixed well.
5. Do not use grossly lipemic specimens.

**ASSAY PROCEDURE**

Prior to assay, allow reagents to stand at room temperature. Gently mix all reagents before use.

1. Dilute specimens, to the necessary range with Phosphate Buffer Saline pH 7.0. (Urine samples are normally diluted 1:20 for a methamphetamine cutoff of 500 ng/ml.) The dilution factor and volume added can be adjusted based on the laboratory's cutoff.
2. Add 10 µl. of appropriately diluted calibrators and standards to each well in duplicate.

3. Add 10 µl. of the diluted specimens in duplicate (recommended) to each well.
4. Add 100 µl of the Enzyme Conjugate to each well. Tap the sides of the plate holder to ensure proper mixing.
5. Incubate for 60 minutes at room temperature (18-26° C) preferably in the dark, after addition of enzyme conjugate to the last well.
6. Wash the wells 6 times with 350 µl. distilled water using either a suitable plate washer or wash bottle taking care not to cross contaminate wells. If testing samples containing abnormally high amounts of hemoglobin (some Postmortem samples), use 10 mM Phosphate buffered saline pH 7.0-7.4. This will lower potential nonspecific binding of hemoglobin to the well, thus lowering background color.
7. Invert wells and vigorously slap dry on absorbent paper to ensure all residual moisture is removed. This step is critical to ensure that residual enzyme conjugate, does not skew results. If using an automated system, ensure that the final aspiration on the wash cycle aspirates from either side of the well.
8. Add 100 µl. of Substrate reagent to each well and tap sides of plate holder to ensure proper mixing.
9. Incubate for 30 minutes at room temperature, preferably in the dark.
10. Add 100 µl. of Stop Solution to each well, to change the blue color to yellow.
11. Measure the absorbance at a dual wavelength of 450 nm and 650 nm.
12. Wells should be read within 1 hour of yellow color development.
13. The following data represent a typical dose/response curve.

**Example of a Standard Curve**

d-methamphetamine ng/ml	OD 450nm
0	1.519
10	0.649
25	0.471
50	0.359

The dose/response curve shown above should not be used in assay calculations. It is recommended that at least one in-house positive quality control sample be included with every assay run. A dose response curve or a cutoff calibrator should be run with every plate.

**RESULTS**

If the average sample absorbance is equal to or less than the average absorbance of the laboratory positive reference standard the sample is POSITIVE for methamphetamine. If the average sample absorbance is greater than the average absorbance of the laboratory positive reference standard the sample is called NEGATIVE for methamphetamine.

Alternatively a dose response curve can be established by plotting standard concentration (abscissa) against corresponding absorbance (ordinate). Values for unknown samples are obtained by interpolation from the curve.